

# #294. Preclinical Assessment of Anti-Tumor Activity and Immune Response in Syngeneic Tumor Models

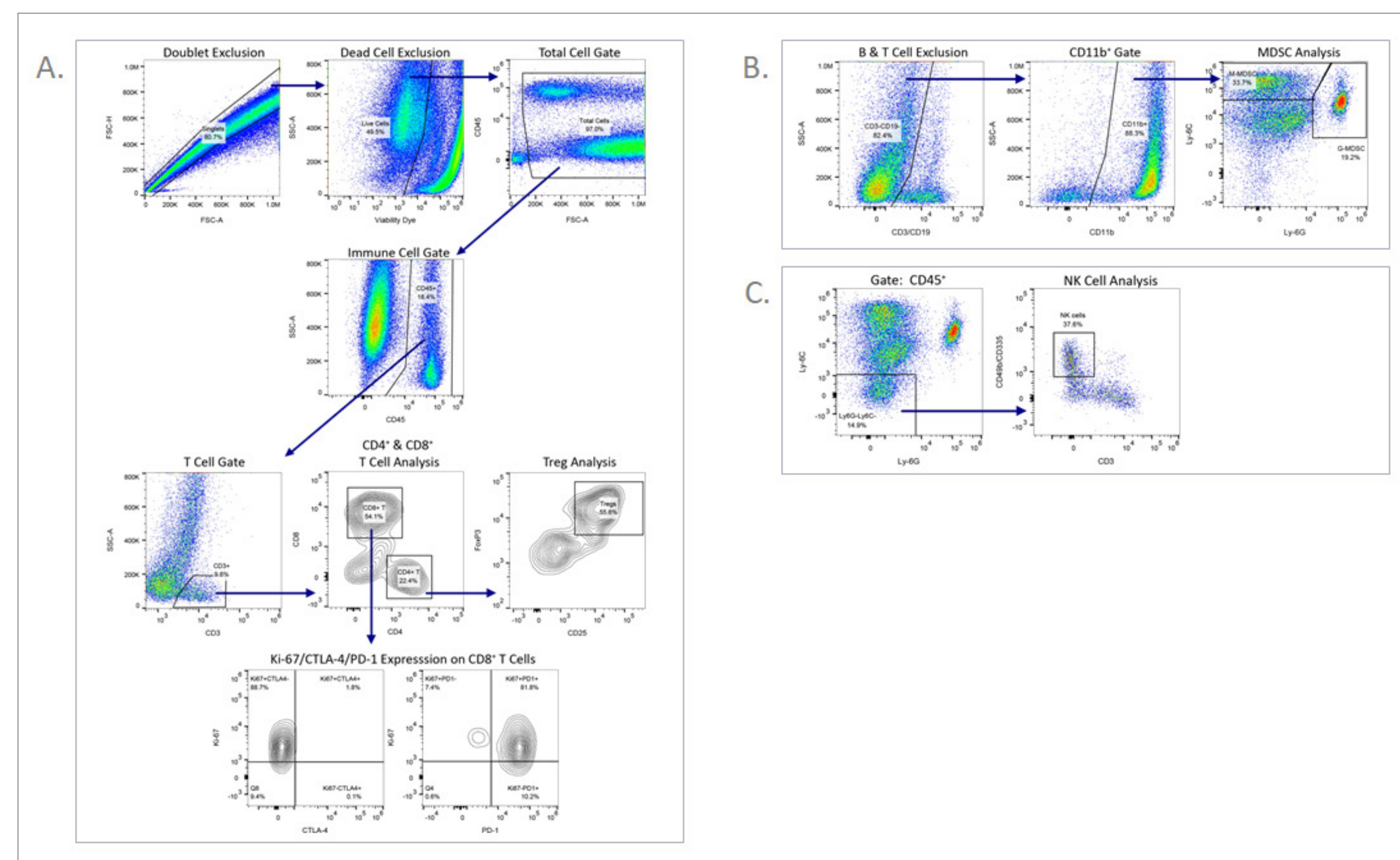
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## Introduction and Background

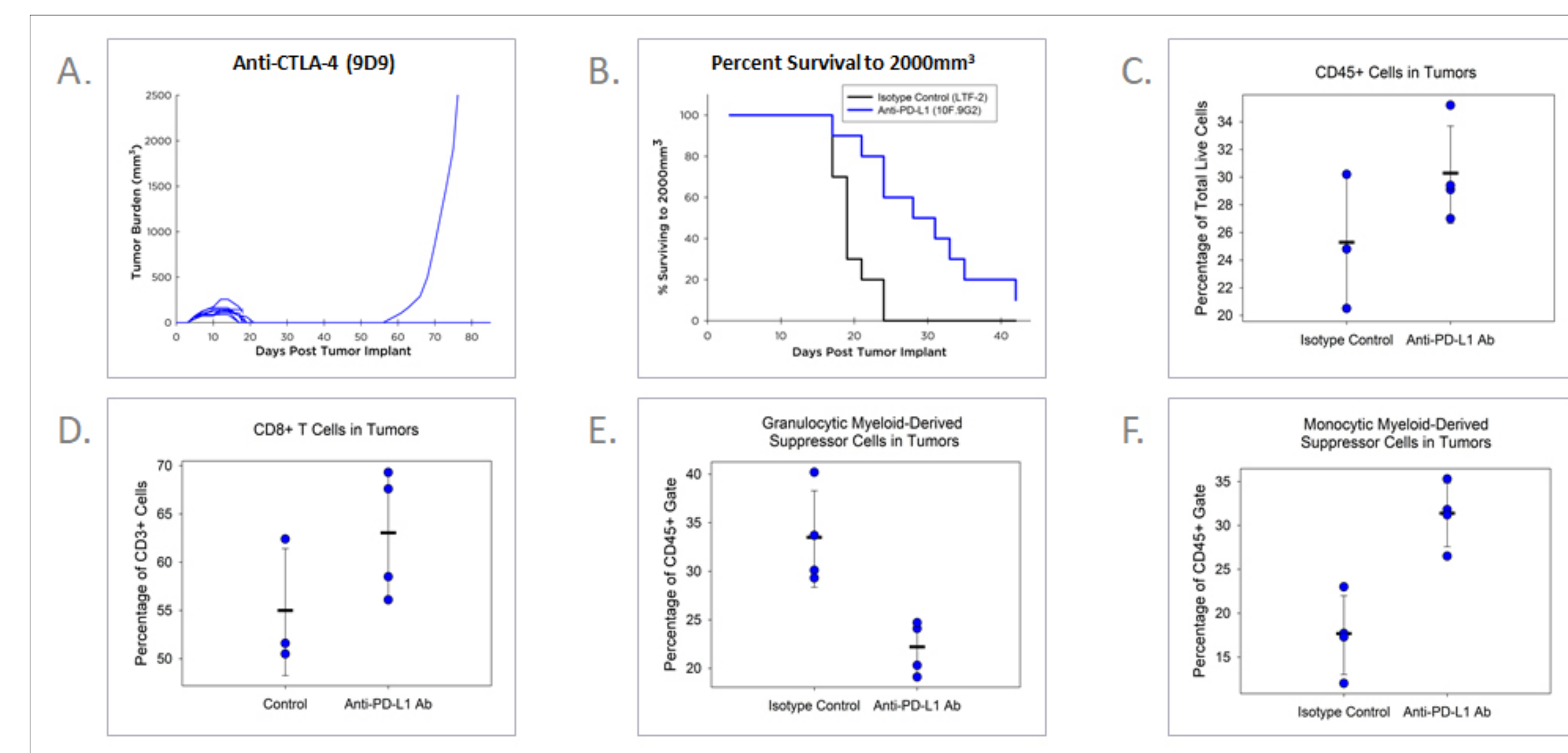
- Preclinical immuno-oncology (I/O) needs identification and refinement of tumor models that recapitulate relevant biological dynamics.
- We tested several murine models for their response to checkpoint inhibitors like anti-CTLA-4, anti-PD-L1 and anti-PD-1 antibodies and found sensitive, moderately sensitive and insensitive models.
- Since the application of more sophisticated endpoints is critical to confidently assess drug sensitivities we also evaluated the immune profiles of these models following treatment.

## Materials and Methods

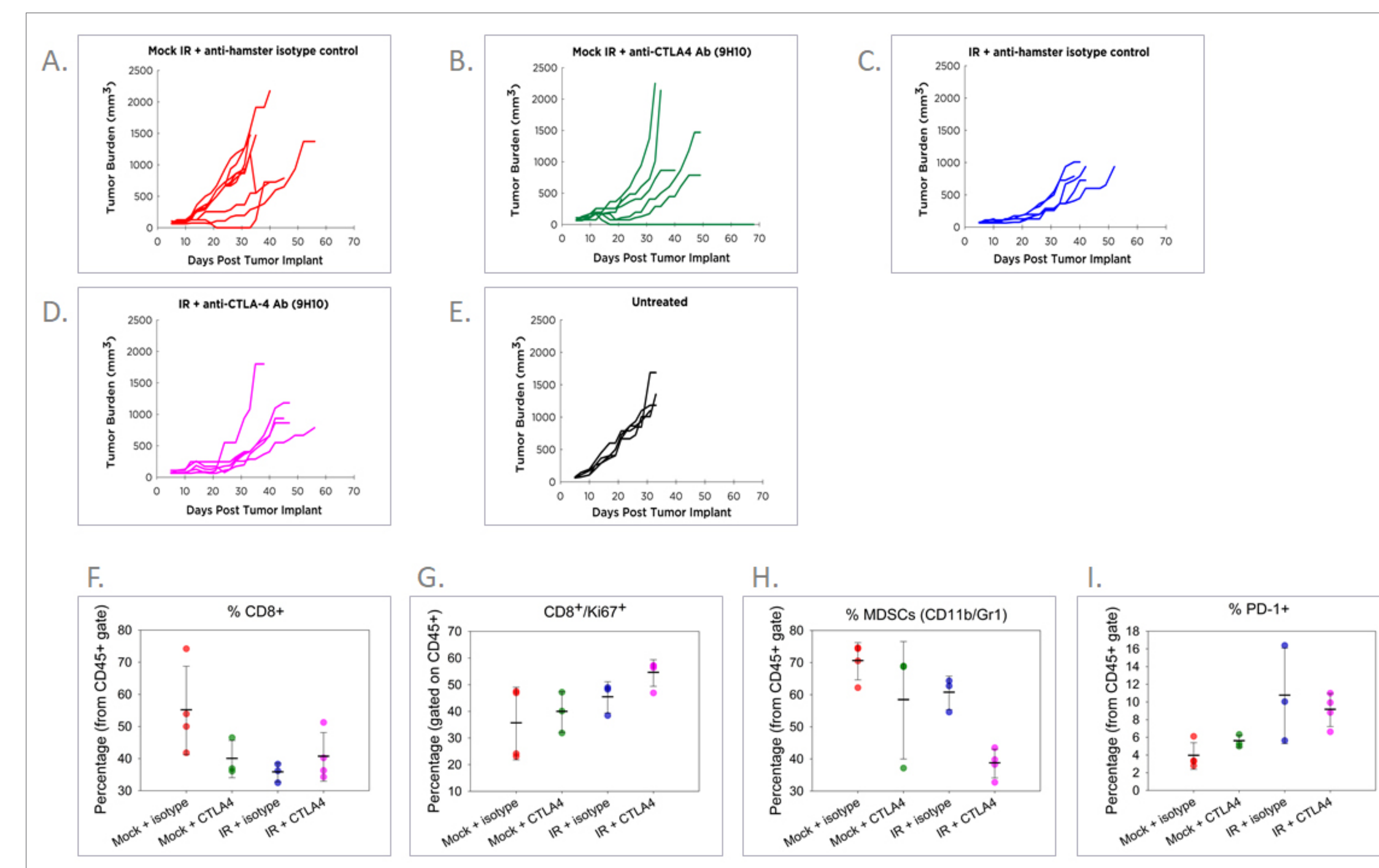
- Female Balb/C mice (CT26, 4T1-Luc) or C57BL/6 mice (Pan02) were purchased from Envigo and were implanted SC in the high axilla (CT26, Pan02) or in the mammary fat pad (4T1-Luc).
- Mice were treated IP with In Vivo Plus antibodies from Bio X Cell (West Lebanon, NH) at 10 mg/kg two times/week for a total of four or five doses.
- In the 4T1-Luc model, localized radiation of 8 Gy at a rate of 1.50 Gy/min was delivered to the tumor area with an RS2000 Biological X-ray Irradiator (Rad Source Technologies, Alpharetta, GA).
- For flow cytometry, the tumors were processed into single-cell suspensions using the gentleMACS™ Dissociators (Miltenyi Biotec). Samples were acquired on an Attune™ NxT Flow Cytometer (Thermo Fisher Scientific) and data was analyzed using FlowJo software (Tree Star).



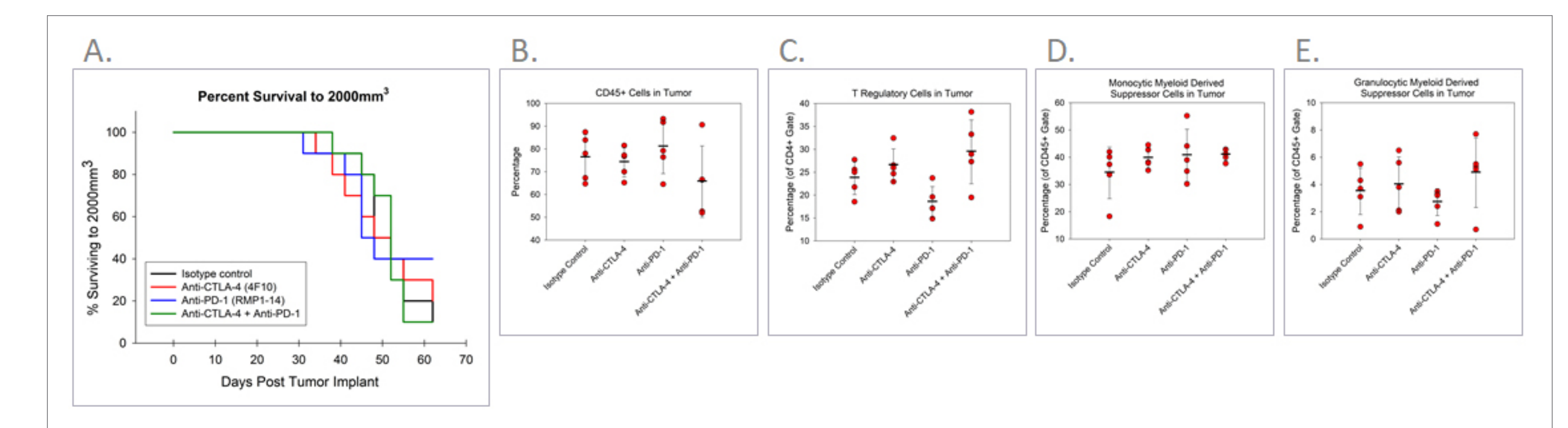
**Figure 1. Gating strategies for flow cytometry.** Similar gating strategies were used to analyze the 4T1 and Pan02 models. A) T cell analysis in CT26 tumors. Following exclusion of doublets and dead cells, total cells were analyzed for CD45+ immune cells. The CD3+ gate was then subdivided into CD4+ and CD8+ T cells. CD4+ T cells were further analyzed for the regulatory T cell subset (Tregs). Finally, CD8+ T cells were further analyzed for the Ki-67 proliferation marker and the CTLA-4/PD-1 exhaustion markers. B) MDSC analysis in CT26 tumors. B and T cells were first excluded from the CD45+ gate. CD11b+ cells were then further analyzed for M-MDSC and G-MDSC subsets. C) NK cell analysis in CT26 tumors. After exclusion of various myeloid subsets using Ly-6G and Ly-6C, NK cells were identified as CD3-CD49b+CD335+.



**Figure 2. CT26: A model sensitive to checkpoint inhibitors (CPIs).** Treatment effects of anti-CTLA-4 (A) or anti-PD-L1 (B) antibody in the CT26 mouse colon carcinoma model. Increased CD45+ cells (C) and CD8+ T cells (D) following treatment with anti-PD-L1 antibody. Treatment with anti-PD-L1 antibody shifts MDSC population from more granulocytic (E) to more monocytic (F).



**Figure 3. 4T1-Luc: A model moderately sensitive to CPIs.** Treatment effects of an anti-CTLA-4 antibody, localized radiation (IR) or the combination in the 4T1-luc mouse mammary carcinoma model. Individual growth over time following treatment with isotype control (A), anti-CTLA-4 antibody (B), radiation (C) or the combination (D). Combination treatment triggers both pro- and anti-tumor signaling pathways thus providing a possible explanation for the marginal anti-tumor responses we observed in this model. Use of precise focal radiation could provide improvements in either single agent IR or IR combined with checkpoint inhibitors.



**Figure 4. Pan02: A model insensitive to CPIs.** Treatment with anti-CTLA-4 or anti-PD-1 antibodies as single agents or in combination displayed no anti-tumor activity (A). Treatment with CPIs did not substantially modulate the immune profile of the tumors (B – E) providing some possible rationale for the lack of efficacy observed.

**Table 1. Comparison of Immune Profiles**

Model/Treatment	Endpoint							
	CD45+ (%Total Cells)	CD4+ T cells (%CD3+)	CD8+ T cells (%CD3+)	Tregs (%CD4+ T cells)	NK (%CD45+)	G-MDSC (%CD11b)	M-MDSC (%CD11b)	
CT26	Control	25.2 ± 4.9	25.6 ± 5.1	54.8 ± 6.6	47.7 ± 3.1	9.05 ± 3.0	33.3 ± 5.0	17.5 ± 4.5
	CTLA-4	ND	ND	ND	ND	ND	ND	ND
	PD-1	ND	ND	ND	ND	ND	ND	ND
	PD-L1	30.2 ± 3.5	19.7 ± 4.1	62.9 ± 6.6	47.9 ± 11.5	9.3 ± 1.4	22.1 ± 2.8	31.2 ± 3.6
4T1	Control	59.4 ± 8.1	63.6 ± 16.5	19.5 ± 16.6	27.7 ± 10.6	0.5 ± 0.3	54.0 ± 8.6	6.7 ± 2.7
	CTLA-4	75.0 ± 7.0	41.2 ± 24.3	15.5 ± 7.8	10.7 ± 9.6	1.4 ± 1.3	39.9 ± 10.3	13.6 ± 5.4
	PD-1	ND	ND	ND	ND	ND	ND	ND
	PD-L1	ND	ND	ND	ND	ND	ND	ND
Pan02	Control	80.4 ± 8.6	19.4 ± 5.7	46.0 ± 9.7	23.7 ± 3.6	1.1 ± 0.1	3.5 ± 1.7	34.3 ± 9.5
	CTLA-4	77.9 ± 8.2	15.4 ± 1.2	46.9 ± 11.1	26.5 ± 3.6	1.0 ± 0.3	4.0 ± 2.0	39.7 ± 3.8
	PD-1	84.3 ± 8.2	19.6 ± 3.9	47.9 ± 3.6	18.5 ± 3.4	1.1 ± 0.1	2.7 ± 1.0	40.7 ± 9.6
	PD-L1	ND	ND	ND	ND	ND	ND	ND

## Results and Conclusions

- The CT26 model is sensitive to immune CPIs with 100% of the mice showing anti-tumor response following treatment with anti-CTLA-4 antibody and 40% demonstrating response following treatment with anti-PD-L1 antibody.
- Treatment of CT26 tumor-bearing mice with anti-PD-L1 results in an increase of CD45+ lymphocytes and modifies the composition of the myeloid derived suppressor cell population.
- Treatment of 4T1-Luc mice with radiation and anti-CTLA-4 antibody triggers both pro- and anti-tumor signaling pathways thus providing a possible explanation for the marginal anti-tumor responses we observed in this model.
- Pan02 is non-immunogenic, similar to human pancreatic cancers. No treatments had anti-tumor effects. The treatments did not alter the immune phenotype of this model. Pan02 may be useful to test CPIs in combination with other I/O agents, targeted agents, chemotherapies or radiation.